Usefulness of Blood Parameters for Preliminary Diagnosis of Brucellosis

Alisha Aky a, Arezoo Bozorgomid a, Kayghobad Ghadiri a, Mahnaz Ahmadi b, Azam Elahi a, Hadi Mozafari c, Afshin Almasi d, Parvin Namadi e, Roya Chegenelorestani a

1Infectious Diseases Research Center, Kermanshah University of Medical Sciences, Kermanshah, Iran; 2Clinical Research Development Center, Imam Reza Hospital, Kermanshah University of Medical Sciences, Kermanshah, Iran; 3Fertility and Infertility Research Center, Health Technology Institute, Kermanshah University of Medical Sciences, Kermanshah, Iran; 4Cardiovascular Research Center, Kermanshah University of Medical Sciences, Kermanshah, Iran

Background: Human brucellosis is a multisystem disease with a wide range of clinical signs which often leads to misdiagnosis and treatment delay. Early diagnosis of this disease can prevent the serious complications and mismanagements. This study aimed to evaluate the hematological parameters with predictive value for the diagnosis of brucellosis.

Methods: In this prospective case-control study which was done during 2015-2017 in Imam Reza Hospital, Kermanshah Province, west Iran, 100 patients with a confirmed diagnosis of brucellosis (brucellosis group) and 100 healthy individuals (control group) were studied. The hematological parameters, including hemoglobin (Hb), red blood cell (RBC), white blood cell (WBC) count, lymphocyte count, neutrophil count, platelet count (PLTs), mean platelet volume (MPV), platelet distribution width (PDW), erythrocyte sedimentation rate (ESR), and C-reactive protein (CRP) of both groups were recorded. The data were statistically compared between the brucellosis and the control groups.

Results: The mean age of patients and healthy groups was 44.04 ± 23.11 and 37.92 ± 24.80, respectively (P = 0.062). The WBC, CRP and neutrophil counts were significantly higher in the brucellosis group (P < 0.05). Based on the receiver operating characteristic (ROC) analysis, the sensitivity and specificity were 54% and 66% for the WBC, 45% and 71% for the neutrophil and 65% and 72% for the CRP, respectively. There was no statistically significant difference between the two groups in terms of Hb, RBC, WBC, lymphocyte and platelet count, MPV, PDW and ESR (P > 0.05).

Conclusion: The results of this study indicate that WBC, CRP and neutrophil count can be used as valuable markers in the preliminary diagnosis of brucellosis. However, further researches are required to standardize these parameters for various forms of brucellosis.

Keywords: brucellosis, diagnosis, blood cell count, blood parameters

Introduction

Brucellosis is an infectious disease caused by a gram-negative coccobacillus of the genus Brucella, with various clinical manifestations. More than half a million people are diagnosed with brucellosis every year.1 After acquiring Brucella, bacteria spread through hematogenous dissemination and affect various organs, such as the urinary, respiratory, central nervous system and cardiovascular. The spread of infection triggers common symptoms of illness, such as fever, nocturnal hyperhidrosis, weight loss, anorexia, arthralgia, and fatigue.1,2 This zoonotic disease, which remains a major public and economic health issue in many developing countries, is endemic to the Middle East, South and Central America, the Mediterranean region, and India.3 Based on reports Iran ranks second in the world in terms of brucellosis and its annual incidence is 98 to 130 people per 100,000 populations.4 Early and
accurate diagnosis of this disease, therefore, plays an important role in controlling and eradicating brucellosis for improving public health. Various laboratory tests, such as bacteriological, serological and molecular methods, have been developed to diagnose brucellosis. While bacterial culture is the gold standard for brucellosis diagnosis, in most cases Brucella culture is not promising due to the lack of optimum conditions. Therefore, serological tests are often used as diagnostic and screening tools. Serological tests sometimes have false results, in particular in case of cross-reactions with other gram-negative bacteria such as Escherichia coli, Yersinia enterocolitica, and Salmonella Urbana. Since a high rate of false-positive results with serological tests, the confirmatory tests are required for positive samples.

Recently, diverse hematological and inflammatory factors have been widely considered as markers of bacterial infections with abundant evidence to support their usefulness in the preliminary diagnosis of infections. Because Brucella is an intracellular bacterium, it can live in phagocytic cells, such as neutrophils and macrophages. Brucellosis is often presented with inflammatory symptoms. Following infection, Brucella spreads to the lymph nodes and from there to the blood and causes systemic infection. As a result, the increase in the number of leukocytes and neutrophils, as well as changes in inflammatory indices, occurs during infection. Research has shown that platelets also contribute to the inflammatory response. Changes in hematological markers are commonly observed in brucellosis. Hematological markers, including white blood cell count, platelet count (PLT), mean platelet volume (MPV), platelet distribution width (PDW), red cell distribution width (RDW), neutrophil to lymphocyte ratio (NLR), platelet to lymphocyte ratio (PLR), monocyte to lymphocyte ratio (MLR), and CRP test, have been used in the preliminary diagnosis of brucellosis, along with serological tests. Both diagnosis and follow-up of treatment for brucellosis are complicated, and it may be helpful to use routine laboratory tests for better management of this disease. The purpose of this research was to evaluate the hematological and inflammatory markers for laboratory diagnosis and follow-up of brucellosis.

Patients and Methods

Participants

This prospective and case–control study was carried out on the brucellosis patients who had been referred to the Imam Reza Hospital between July 2015 and March 2017. This Hospital is the referral center for infectious diseases located in Kermanshah, West of Iran. Furthermore, as a control group, the healthy people of similar age and gender admitted to the Imam Reza Hospital for routine check-up were selected during the same period.

The diagnosis of brucellosis cases was based on clinical of symptoms (fever, joint pain, sweating and fatigue) and laboratory results (Wright, Coombs Wright and 2-mercaptoethanol (2ME) tests). People with a history of brucellosis or inflammatory diseases, anemia, malignancies, platelet disorders, blood transfusion within the last 3 months, HIV infection, diabetes mellitus, hypertension and other kinds of diseases were excluded from this study. None of the subjects had received steroid therapy or another anti-inflammatory drug. Information regarding age, sex and medical history were recorded from individual’s files or interviews. All subjects were agreed to sign an informed written consent for this study. This study was approved by the Ethics Committee of the Kermanshah University of Medical Sciences.

Blood Test

Five mL of venous blood sample was collected from the participant and stored in tubes containing EDTA. A complete blood count analysis was done with automated analyzers Coulter HmX from Beckman Coulter at admission. Hemoglobin (Hb), Red blood cell (RBC) count, white blood cell (WBC) count, lymphocyte count, neutrophil count, platelet count (PLTs), mean platelet volume (MPV), platelet distribution width (PDW), erythrocyte sedimentation rate (ESR), and C-reactive protein (CRP) were recorded for each individual. The reference values in the affiliated Imam Reza laboratory were 3.5–9.5 ×10^9/L for WBC count, 1.8–6.3 ×10^9/μL for neutrophil count, 40–70% for neutrophil percentage, 181–300 ×10^9/μL for PLT, 9.4–12.5 fL for MPV and 15.5–18.1% for PDW.

Data Analysis

The SPSS software was used for statistical analysis (version 20, SPSS Inc., Chicago, IL, USA). The Kolmogorov–Smirnov test was applied to evaluate the normal distribution of the data in each group. Continuous variables have been presented as mean ± standard deviation (SD). Independent samples t-test or Mann–Whitney U-test was used for comparison of two groups. Categorical variables were compared using chi-square test. The cut-off value for WBC and neutrophil count that best distinguishes between
healthy controls and brucellosis patients were performed using receiver operating characteristic (ROC) curves analysis, for which sensitivity and specificity values were calculated. Correlations between numerical variables were assessed using Pearson’s or Spearman correlation analysis. P value <0.05 was taken as significant.

**Results**

The 100 brucellosis patients and 100 healthy people with age and gender matched as the control group were included in this study. There were 52 male participants in the brucellosis and 59 in the control group (P =0.319). Mean age of brucellosis and the control group was 44.04 ± 23.11 and 37.92 ± 24.80 years, respectively (P =0.062). There was no statistical difference between the two groups in terms of age and sex (P>0.05).

The comparisons of blood parameters between brucellosis and control groups have been given in Table 1. No significant difference was detected in the RBC count, neutrophil count, lymph count, ESR percentage, Hb, RDW, PLRC, PDW, PLT, and MPV values between the two groups (P>0.05). The WBC and neutrophil counts were 7.1 and 57 in the control group, which were significantly lower than that in brucellosis patients with 8.9, and 65, respectively (P < 0.05).

Based on the ROC analysis, in the brucellosis group, the cut-off values of the WBC and neutrophil were 7.55 and 53.7, respectively (Figure 1). The sensitivity and specificity were 54% and 66% for the WBC, 45% and 71% respectively.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Statistic Information</th>
<th>Brucellosis (n = 100)</th>
<th>Controls (n = 100)</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>Mean±SD</td>
<td>44.04 ± 23.11</td>
<td>37.92 ± 24.80</td>
<td>0.062*</td>
</tr>
<tr>
<td></td>
<td>Range</td>
<td>1–84</td>
<td>1–92</td>
<td></td>
</tr>
<tr>
<td>Sex</td>
<td>Male (%)</td>
<td>52</td>
<td>59</td>
<td>0.319 b</td>
</tr>
<tr>
<td>WBC/mm³</td>
<td>Mean±SD</td>
<td>8.92 ±5.59</td>
<td>7.13±3.61</td>
<td>0.004*</td>
</tr>
<tr>
<td></td>
<td>Range</td>
<td>2.8–29.7</td>
<td>2.8–45.6</td>
<td></td>
</tr>
<tr>
<td>RBC/mm³</td>
<td>Mean±SD</td>
<td>4.42±0.65</td>
<td>4.46±0.73</td>
<td>0.680*</td>
</tr>
<tr>
<td></td>
<td>Range</td>
<td>2.8–6.1</td>
<td>2.2–6.7</td>
<td></td>
</tr>
<tr>
<td>ESR, mm/hr</td>
<td>Mean±SD</td>
<td>32.09±21.79</td>
<td>29.02±25.77</td>
<td>0.059*</td>
</tr>
<tr>
<td></td>
<td>Range</td>
<td>2–125</td>
<td>2–125</td>
<td></td>
</tr>
<tr>
<td>Hb, g/dL</td>
<td>Mean±SD</td>
<td>12.2±1.96</td>
<td>12.4±2.39</td>
<td>0.410*</td>
</tr>
<tr>
<td></td>
<td>Range</td>
<td>10.4–13.9</td>
<td>10.8–14.5</td>
<td></td>
</tr>
<tr>
<td>PLT/mm³</td>
<td>Mean±SD</td>
<td>244.33±91.26</td>
<td>251.27±132.60</td>
<td>0.555*</td>
</tr>
<tr>
<td></td>
<td>Range</td>
<td>36–606</td>
<td>44–863</td>
<td></td>
</tr>
<tr>
<td>MPV, fl</td>
<td>Mean±SD</td>
<td>9.08±1.25</td>
<td>9.31±1.09</td>
<td>0.182*</td>
</tr>
<tr>
<td></td>
<td>Range</td>
<td>6.7–12.5</td>
<td>6.8–12.5</td>
<td></td>
</tr>
<tr>
<td>PDW, %</td>
<td>Mean±SD</td>
<td>10.86±2.21</td>
<td>11.18±2.11</td>
<td>0.069*</td>
</tr>
<tr>
<td></td>
<td>Range</td>
<td>8.1–20.7</td>
<td>7.7–19.9</td>
<td></td>
</tr>
<tr>
<td>PLRC, %</td>
<td>Mean±SD</td>
<td>19.69±7.85</td>
<td>20.55±7.24</td>
<td>0.142*</td>
</tr>
<tr>
<td></td>
<td>Range</td>
<td>5.4–51.4</td>
<td>5.3–42.9</td>
<td></td>
</tr>
<tr>
<td>RDW, %</td>
<td>Mean±SD</td>
<td>14.19±1.56</td>
<td>13.96±1.87</td>
<td>0.103*</td>
</tr>
<tr>
<td></td>
<td>Range</td>
<td>11.5–20.2</td>
<td>11.2–23.4</td>
<td></td>
</tr>
<tr>
<td>Neut./mm³</td>
<td>Mean±SD</td>
<td>65.23±16.93</td>
<td>57.94±13.99</td>
<td>0.001*</td>
</tr>
<tr>
<td></td>
<td>Range</td>
<td>8–97</td>
<td>20–89</td>
<td></td>
</tr>
<tr>
<td>Lymph./mm³</td>
<td>Mean±SD</td>
<td>25.43±11.07</td>
<td>29.02±13.50</td>
<td>0.064*</td>
</tr>
<tr>
<td></td>
<td>Range</td>
<td>1–89</td>
<td>6–73</td>
<td></td>
</tr>
<tr>
<td>CRP</td>
<td>Positive (%)</td>
<td>65</td>
<td>28</td>
<td>&lt;0.001 b</td>
</tr>
</tbody>
</table>

Notes: aMann–Whitney U-test, bChi-square test, cIndependent sample t-test.
Abbreviations: WBC, white blood cell; Hb, Hemoglobin; RBC, Red blood cell; NLR, neutrophil to lymphocyte ratio; PLR, platelet to lymphocyte ratio; MPV, mean platelet volume; ESR, erythrocyte sedimentation rate; CRP, C-reactive protein; PDW, platelet distribution width.
for the neutrophil and 65% and 72% for the CRP, respectively. The area under the curve (AUC) of the WBC and neutrophil were 0.62 and 0.58, respectively (Table 2).

Spearman correlation coefficient revealed an inverse and significant correlation between the number of neutrophils and the number of lymphocytes (r coefficient: -0.817, P<0.001). In other words, as the number of neutrophils increased, the number of lymphocytes decreased. Also, the number of neutrophils showed a direct and significant relationship with the number of WBC (r coefficient: -0.817, P<0.032).

Discussion

Brucellosis is an important health problem in Iran, where animal husbandry is an important livelihood. According to a systematic review and meta-analysis in Iran, the highest rate of brucellosis was related to Kermanshah Province with 276.42/100,000 people.\(^{16}\) \(B.\) \textit{mellitensis} accounts for the majority of the disease in humans in various provinces of Iran, followed by \(B.\) \textit{abortus}.\(^{17}\)

This is not the first study on changes in the hematological parameters in patients with brucellosis. However, given the impact of biological and environmental factors on microbial infection, it can provide a better local view for these parameters. Research results show that hematological parameters in patients with brucellosis differ from patient to patient and these parameters can return to their normal value after the treatment of brucellosis.\(^6\) The inflammatory process that occurs during brucellosis is associated with an increase in acute-phase reactants.\(^18\)

CRP is an acute-phase protein that increases up to 1000-fold in the blood of patients with infection or inflammation. During some bacterial infections, CRP level elevates in the response to cytokines, mainly tumor necrosis factor-\(\alpha\), interleukin (IL)-1\(\beta\) and IL-6.\(^19\) The production of this cytokine is one of the primary phagocyte responses.\(^19\) CRP is able to binds to various bacteria with exposed phosphocholine (PCh) groups. This interaction activates the classical complement system to destroy the ligand and kill off the pathogen.\(^20\) However, Healy and Freedman suggested that the level of serum CPR could only indicate the presence of infection.\(^21\) A study in Turkey reported that the level of CRP was higher in osteoarticular brucellosis patients than in non-osteoarticular brucellosis patients.\(^22\) Another study reported that patients with the acute brucellosis exhibited higher CRP levels than the control group.\(^6,13,23\) Some other studies have suggested that elevated CRP level may also be associated with disease severity and mortality in hospitalized patients with community-acquired pneumonia.\(^24\) In our study, CRP was significantly higher in brucellosis patients, which indicates that CRP is a valuable marker for the diagnosis of brucellosis.

WBC count can be considered as a marker for leukocytosis in brucellosis.\(^13\) Neutrophils and lymphocytes play an important role in inflammatory processes. Physiological immune responses are characterized by an increase in the number of neutrophils and a decrease in the number of lymphocytes.\(^6\) Our findings revealed that the mean of WBC and neutrophil count in \(Brucella\) patients were significantly higher than in control groups. Therefore, they can be reliable markers for the evaluation of brucellosis along with other clinical findings and blood marker changes. Research has shown that the number of leukocytes and neutrophils increases in infections, especially in bacterial infections that indicate an inflammatory response.\(^25,26\)

### Table 2 Results of ROC Analysis for Blood Parameters

<table>
<thead>
<tr>
<th>Parameter</th>
<th>AUC</th>
<th>Cut Off Point</th>
<th>Sensitivity (%)</th>
<th>Specificity (%)</th>
<th>+PV</th>
<th>-PV</th>
<th>+LR</th>
<th>-LR</th>
<th>Accuracy</th>
</tr>
</thead>
<tbody>
<tr>
<td>WBC count (10(^9)/μL)</td>
<td>0.62</td>
<td>7.55</td>
<td>54</td>
<td>66</td>
<td>0.61</td>
<td>0.58</td>
<td>1.58</td>
<td>0.696</td>
<td>0.60</td>
</tr>
<tr>
<td>Neutrophil count</td>
<td>0.58</td>
<td>53.7</td>
<td>45</td>
<td>71</td>
<td>0.60</td>
<td>0.56</td>
<td>1.55</td>
<td>0.774</td>
<td>0.58</td>
</tr>
<tr>
<td>CRP (ng/L)</td>
<td></td>
<td>–</td>
<td>65</td>
<td>72</td>
<td>0.69</td>
<td>0.67</td>
<td>2.32</td>
<td>0.486</td>
<td>0.68</td>
</tr>
</tbody>
</table>

**Abbreviations:** WBC, white blood cell; CRP, C-reactive protein; AUC, areas under the ROC curves; LR, likelihood ratio; PV, predictive value.
Brucella lipoproteins also have pro-inflammatory properties through direct neutrophil activation. The results of the study by Aktar et al showed that the mean values for neutrophils and leukocytes in children with Brucella arthritis were higher than the control group (P<0.05). Imani-Rastabi et al examined changes in blood factors in sepsis and indicated that there was a statistically significant difference in the WBCs count before and after sepsis.

In addition to regulating the immune system responses, studies have reported that platelets also play an active role in the inflammation process. For this purpose, when platelets are activated, they actively participate in host defense through phagocytosis and the development of cytotoxic-free radicals and oxidative molecules. Platelets are involved in inflammatory responses through the involvement of neutrophils and macrophages, increased vascular permeability, leukocyte infiltration and inflammatory mediators such as cytokines and chemokines. It has been reported that in thrombocytopenia, edema and leukocyte infiltration are reduced. MPV is an essential platelet marker linked to the platelet activity and function. PDW which indicates changes in platelet size is correlated with the activity of the platelet. Changes in this index have been reported as a marker to evaluate various inflammatory and infectious diseases. In our study, although the mean values for PLT, MPV, and PDW were lower in the brucellosis group, there were no statistically significant changes (P>0.05). Our findings are consistent with the Togan et al, which found that MPV levels were in the normal range in treatment and control groups. Several studies suggested that the values of MPV and PLT were lower in the brucellosis group than in the control group (P<0.05). Nevertheless, a number of studies have documented that differed MPV levels in specific inflammatory conditions. One indicated that in children with Kawasaki disease, MPV was not a valuable marker for predicting coronary artery abnormalities. A second reported that the benefit of MPV as an inflammatory marker to determine the disease activity in TB patients and a third noted a higher MPV in children infected with Helicobacter pylori infection than in healthy controls. Overall, it is thought that during an acute infection such as septicemia the initial increase in MPV level can be related to thrombocytopenia and during chronic or persistent bacterial infection a delayed decline in MPV level can be related to thrombocytosis.

Here, we should report some limitations of this study. The sample size was small and failed to the evaluation of all clinical forms of brucellosis such as acute, chronic, or focal forms. Therefore, to reach more valuable results, a more comprehensive study in several centers with a larger sample size is recommended.

Conclusion
As a result, hematological parameters in patients with brucellosis are relatively different in various studies. Our analysis and most of the studies have shown that the most significant signs of brucellosis are an increase in the number of leukocytes and a decrease in the number of thrombocytes and lymphocytes. It seems that along with clinical symptoms and serological and culture methods, hematological parameters, such as WBC, neutrophil, and CRP which are inexpensive and available in hospitals, can be useful in the preliminary diagnosis and assessment of brucellosis infection.

Data Sharing Statement
The data sets used and/or analyzed during this study are available from the corresponding author on reasonable request and were received permission for use by the Kermanshah University of Medical Sciences Ethics Committee.

Ethics and Consent Statement
All the participants provided written informed consent prior to the study and this study was conducted in accordance with the Declaration of Helsinki. The protocol was approved by the Kermanshah University of Medical Sciences Ethics Committee.

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Disclosure
The authors report no conflicts of interest in this work.

References


